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# Soil biotic and abiotic conditions negate invasive species performance in native habitat

Pushpa G. Soti<sup>1\*</sup>, Matthew Purcell<sup>2</sup> and Krish Jayachandran<sup>3</sup>

## Abstract

**Background:** Most studies on plant invasion consider the enemy release hypothesis when analyzing native habitats. However, the lower performance of invasive species in the native habitats can be the result of unfavorable soil conditions in the native habitats. While soil biotic and abiotic factors have a potential to restrict the growth of invasive species in their native habitats, our understanding of belowground environment of invasive species in their native habitats is very limited. In this study, we analyzed soil characteristics associated with an exotic invasive plant, Old World Climbing Fern (*Lygodium microphyllum*), in its native habitat in Australia and the recipient habitat in South Florida. Rhizosphere soil samples from both habitats were analyzed for soil physical, chemical and biological characteristics.

**Results:** Soil characteristics in the recipient habitats were significantly different compared to those in the native habitats. Soil samples from the native habitat had low soil pH, and high concentrations of elements such as aluminum and zinc which are phytotoxic in acidic soil environments. Additionally, mycorrhizal fungi spores were more diverse in the recipient habitat in Florida compared to the native habitat in Australia.

**Conclusion:** Overall, our results indicate that growth of an invasive plant in its native habitats could be restricted by the toxic effects associated with strong soil acidity. Results from this study indicate that invasive plants not only escape from their natural herbivores but also from toxic soil environment in their native habitats.

**Keywords:** Soil toxicity, Soil chemistry, Soil microbes, Exotic invasive species, *Lygodium microphyllum*

## Background

Biotic and abiotic interactions of exotic invasive plants at both below and above ground can vary significantly in the native and recipient habitats. The potential of any exotic species to invade a new habitat generally depends on abiotic tolerance (Alpert et al. 2000; Levine et al. 2004), biotic resistance (Case 1990; Levine et al. 2004; Bogdziewicz et al. 2019), and propagule pressure (Lockwood et al. 2005; Simberloff 2009), or the combination of these three factors (Byun et al. 2018). Most research on exotic invasive species is focused on plant traits such as high growth rate, short lifecycle, high levels of resource allocation to reproduction, and flexible utilization of available environmental resources. Several studies

with biogeographic comparison of invasive species in their native range and invaded range focus on the impact of variable soil microbes on plant performance (Callaway and Aschehoug 2000; Hierro et al. 2005; Vermeij et al. 2009; Volin et al. 2010). While there is increased research interest in the microbiome of invasive species (Coats and Rumpfo 2014; Kamutando et al. 2019; Ramirez et al. 2019), there is limited information on the performance of invasive plants in their native habitats and the restrictions posed by the soil characteristics. For example, soil pH has complex effect on plant growth leading to the variation in the distribution of plant species in acidic or calcareous soils. Diekmann and Lawesson (1999) reported that pH is one of the major underlying variables determining the floristic variation within forest communities. In addition, soil acidity can result in toxicities of aluminum and manganese and deficiencies or low availability of certain essential elements

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including calcium, magnesium, phosphorus, and molybdenum (Foy 1984). These factors may directly or indirectly restrict plant growth through interference in the development and functioning of symbiotic associations with rhizobia, mycorrhizas, actinomycetes, and other plant growth promoting rhizobacteria. It is reported that soil acidity as a result of Al toxicity has detrimental effects on plant growth by lowering rooting depth, increasing susceptibility to drought, and decreasing uptake of subsoil nutrients. Plants exposed to Al toxicity are reported to have stunted growth, small dark green leaves, late maturity, and thick root tips and lateral roots. They have many stubby lateral roots but lack in fine branching, thus, inefficient in absorbing nutrients and water (Kochian et al. 2004). Similarly, soil texture is also an important factor influencing the distribution of minerals, organic matter, microbial community, and other soil properties (Scott and Naiman 2006).

Once established, invasive species create a favorable rhizosphere conditions in the recipient habitats (Ehrenfeld 2003; Perkins et al. 2011; Coats and Rumpfo 2014) and increase their fitness (Soti et al. 2019). While it is generally unlikely to eliminate invasive plant species, the recipient habitats can probably be managed to maximize the performance of native species relative to that of invaders by altering resource levels and disturbance regimes (Daehler 2003). Information on the soil conditions, where the plant has adapted, can provide a key insight in integrating environmental manipulations into the habitat restoration projects, but the comparative analysis soil characteristics in the native habitat of an invasive species remain largely unexplored.

Here, we analyze the role of soil chemistry in the successful invasion of south Florida's natural communities by a highly invasive plant species, *Lygodium microphyllum* (Old World climbing fern). *L. microphyllum* is an invasive exotic plant species taking over many sites in freshwater and moist habitats across southern and central Florida. In the invaded regions of South Florida *L. microphyllum* displays most of the ecological characteristic associated with successful invasive plants (Westbrooks 1998): it has the ability to grow in varying hydrological (Gandiaga et al. 2009), nutrient (Volin et al. 2010), and light conditions (Volin et al. 2004). It tolerates a wide range of soil pH (Soti et al. 2014, 2015), and has a strong symbiotic relationship with arbuscular mycorrhizal fungi (AMF) (Soti et al. 2014). Comparative analysis of soil samples from both its native range and invaded region have shown that *L. microphyllum*, which had adapted to close-to-neutral soils in Florida, grows in highly acidic soils in its native range in Australia (Soti et al. 2014). Additionally, the Food and Agriculture Organization (FAO) world soil distribution map shows the soil in the native range, identified by Goolsby et al.

(2003), is a region with high soil acidity. Volin et al. (2010) indicated that the growth of *L. microphyllum* was highest in sandy soils which indicates that this plant prefers well drained sandy soils. Furthermore, the root and rhizome growth of *L. microphyllum* was highest in the sandy soils of south Florida compared to the native Australian soils.

*L. microphyllum* does not have a high economic value in its native range, thus there is very little information available about its native ecology, and the available information is mostly on its native herbivores. Very little is known about why this plant is invasive outside its native range (Ferriter 2001). In the invaded regions in South Florida, *L. microphyllum* is highly invasive and current management techniques such as fire, herbicides, and mechanical removal are inadequate. The prospect of developing a method that targets the rhizomes of *L. microphyllum* is compelling; however, the soil biogeochemical characteristics of *L. microphyllum* infested sites remain unexplored. Before we target the rhizomes, it is imperative to elucidate the soil characteristics of the sites, where this plant has adapted, to obtain baseline information. Additionally, it is important to gain information of the soil characteristics of the native range where this plant has its origin and adapted habitat. Results from this study will assist in better understanding of the complex feedbacks between exotic invasive plants, soil microbial community and soil elements.

To determine if *L. microphyllum* escaped from the toxic soil conditions and natural soil enemies, we explore the variation in the biotic and abiotic soil characteristics of *L. microphyllum* from both the native and recipient habitats. We hypothesized that the rhizosphere soil in the native habitat is generally poor in nutrients and high in phytotoxic elements such as Al compared to the recipient habitat in Florida.

## Methods

### Sites selection

We compared soil samples collected from three different sites each in Queensland, Australia, where the plant species is native, and in Florida, United States, where *L. microphyllum* is a Category I exotic invasive ([FLEPPC] Florida Exotic Pest Plant Council 2019). Queensland and South Florida have similar climate conditions such as average monthly maximum and minimum temperatures, rainfall amounts, rainfall pattern, and relative humidity (for details see Goolsby et al. 2003). In its native Australian sites, *L. microphyllum* grows around the edges of *Melaleuca quinquenervia*, a major invasive plant in the South Florida or edges of swamps along with *Drynaria* sp. and *Blechnum indicum*. In the recipient habitat Florida, the plant has adapted to multiple hydrologic conditions. The three study sites in Florida were selected based on the different habitat

conditions: (1) Tree Tops Park, a highly disturbed habitat which is seasonally flooded, with *L. microphyllum* growing in the understory; (2) Jonathan Dickinson State Park, is relatively undisturbed habitat with sand pine scrub communities. Here, *L. microphyllum* grows to the top of the canopy of trees; and (3) Central Florida site, a private property with sand mine spoil dominated by *L. microphyllum* and a few native ferns.

### Sample collection

Soil samples were collected during the dry season in both continents (Table 1, Fig. 1), except for the Central Florida site (which was added later at the request of Nature Conservancy). At each sampling site, six 1 m × 1 m plots were selected randomly, and soil from the 10–15 cm deep zone was collected from all four corners and the center of each plot with a soil corer (diameter: 18 mm) and mixed homogeneously into one bulk sample for each plot, resulting in six composite soil samples from the six different sites in the two habitats. The soil samples from the local sites in South Florida were transported to the laboratory in a cooler, and samples from Australia and Central Florida were stored in 4 °C and shipped overnight.

### Soil nutrient analysis

A small portion of each soil sample was air dried and passed through a 2 mm sieve for analysis of physico-chemical properties. They were ground to fine powder with a mortar and pestle and stored at room temperature in air-tight containers for further analysis of nutrients and trace elements. The soil pH was measured with a pH meter (soil solution ratio 1:2 in water), texture was measured by the hydrometer method (Bouyoucos 1962), and total organic matter was measured based on the standard loss on ignition method (500 °C, 5 h; Storer 1984). Total C and N in soil and leaves were measured with a Truspec CN analyzer. Total Ca, Fe, Al, Mg, K, Mn and P in soil were measured with an ICP–MS at USDA, ARS Laboratory, Miami, Florida after following the acid digestion Method 3050B (USEPA Method 3050 1996). One gram of finely ground soil samples was

transferred to large glass tubes and mixed with 10 ml of 30% HNO<sub>3</sub>. The tubes were covered with a vapor recovery system and heated to 95 ± 5 °C and refluxed for 10 min without boiling under the hood in a heating block maintained with a Partlow Mic 6000 Profile Process Controller.

After cooling to 40 °C, 5 ml of concentrated HNO<sub>3</sub> was added and the samples were then heated at to 95 ± 5 °C until no brown fumes were given off. The samples were again cooled to 40 °C, 2 ml of DI water and 3 ml of 30% H<sub>2</sub>O<sub>2</sub> were added and heated until the effervescence subsided. The samples were cooled and diluted to 50 ml with DI water, centrifuged at 2000 rpm for 10 min and filtered with a Whatman No. 41 filter paper.

### Microbial analysis: bacteria and fungi population

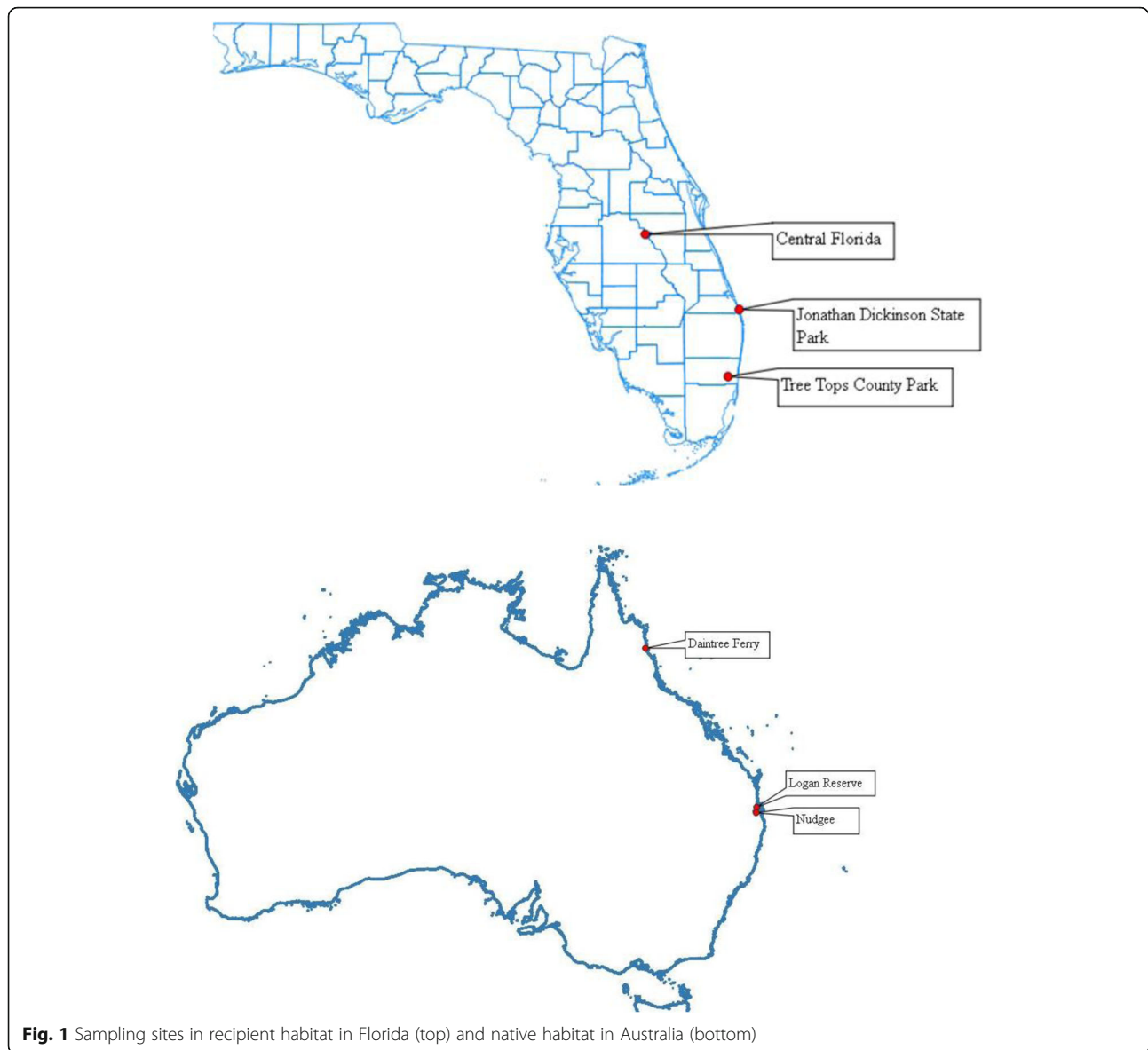
Total colony forming units (CFU) of bacteria and fungi was determined by the standard spread plate dilution method as described by Seeley Jr and VanDemark (1962). Dry equivalent of one-gram soil was mixed in 9 ml sterile water (autoclaved) and was diluted serially. Samples were vigorously mixed during dilution to assist in dislodging the bacteria from the soil particles. A serial dilution of 10<sup>-2</sup>, 10<sup>-3</sup>, 10<sup>-4</sup>, and 10<sup>-5</sup> was made for fungi and 10<sup>-4</sup>, 10<sup>-5</sup>, 10<sup>-6</sup>, and 10<sup>-7</sup> for bacteria. A total of 100 µl of diluted soil suspension was spread on three plates per soil sample for both bacteria and fungi at each dilution level. Nutrient agar containing cycloheximide solution (to prevent fungal growth) was used for bacteria and Rose Bengal Agar (RBA) with streptomycin sulphate (to prevent bacteria growth) was used for the estimation fungal colonization. Agar plants with spread with sterilized water were used as control. Inoculated plates were incubated at 26 °C for 3 days before the colonies were counted. Dilution plates with 100 to 300 colonies per plate were counted.

### Arbuscular mycorrhizal fungi (AMF) spore extraction and identification

Results from our previous study indicated that the mycorrhizal root colonization in *L. microphyllum* is

**Table 1** Sampling sites and dates of sample collection in the recipient habitat and native habitat of *L. microphyllum*

Sites		Coordinates	Sampling dates
<i>Recipient, FL, US</i>			
1	Tree Tops Park	26° 4'0.04"N, 80° 16' 5.88"W	Dec, 2010
2	Central Florida	28° 23' 4.03" N, 81° 44' 41.30" W	June, 2012
3	Jonathan Dickinson	27°0'37.33"N, 80°7'20.28"W	Dec, 2010
<i>Native, Queensland, AU</i>			
1	Daintree Ferry	16°15'25.57"S, 145°24'3.94"E	June, 2011
2	Logan Reserve	27°40'4.16"S, 153°16'0.44"E	June, 2011
3	Nudgee	27°22'31.12"S, 153° 5'39.42"E	June, 2011



**Fig. 1** Sampling sites in recipient habitat in Florida (top) and native habitat in Australia (bottom)

significantly higher in the invaded regions compared to the native regions in Australia (see Soti et al. 2014). In this study, we identified the morphospecies of AMF spores in the rhizosphere soil of *L. microphyllum* in both the regions following the wet sieving technique (Gerde-mann and Nicolson 1963). 100 ml of DI water was added to dry equivalent of 50 g of soil from each site. It was then mixed vigorously to separate the spores from soil aggregates. The mixture was washed through a series of sieves (2 mm, 100  $\mu$ m and 32  $\mu$ m). Washing was done until the water flowing through the sieves was clear. The sievate retained on the sieves was washed and centri-fuged with water to remove floating organic debris and the supernatant was discarded. The pellet in the bottom was re-suspended in a 50% sucrose solution and centri-fuged for one minute at 2000 RPM to separate the

spores from denser soil components. Immediately after centrifugation, spores in the sucrose supernatant were rinsed in a fine sieve to remove the sucrose. The spores were then washed into a filter paper for vacuum filtra-tion. The fungal spores were then mounted on slides for taxonomic identification to the genus level based on the spore morphology and wall characteristics, using the descriptions by INVAM (International culture collection of vesicular-Arbuscular Mycorrhizal Fungi) (n.d.). The genus that was dominant was taken as the representative mycorrhizal AMF type for each site.

#### Statistical analysis

All soil variables were subjected to normality test. The data was not normalized with transformation, so a non-parametric test was conducted. Kruskal-Wallis test was

done to compare the difference in the rhizosphere soil status of *L. microphyllum* among the different sites. Contrasts were done to test the difference among sites with the overall model was significant. Pearson's correlation analysis and was done with all sites pooled to determine relationship between the measured soil variables. Differences are reported as significant for tests with  $P$ -values  $\leq 0.05$ . All the parameters were analyzed with SAS Version 9.2 software.

## Results

### Soil chemical and physical properties

Soil properties in the study sites were highly variable and site specific (Tables 2, 3; Fig. 2). Soil texture, shown in Table 2, varied significantly among the different sites with native sites having heavier soils (high bulk density) compared to the recipient sites. Soil chemistry results also indicated site specific characteristics in the native and recipient habitats (Table 3; Fig. 2). The Australia sites had strong acidic soil ranging from pH = 4.1 to 4.55 while the recipient sites Florida had significantly higher soil pH ranging from 5.60 to 6.57. In general, elements which are phytotoxic in acidic soil such as Al, Zn, and Fe were higher in the native sites in Australia except for site 2, a sand mine spoil, in the recipient habitat (Fig. 2). Total C and N were highest in the recipient site, and there was no difference in P among the sites in both native and recipient habitats.

### Bacteria and fungi populations

The average number of bacteria and fungi, colony forming units (CFU) per gram of dry soil, was significantly different in all six sites (Table 2). Total CFU of bacteria was highest in native habitat ( $288 \times 10^6$ ) and lowest in recipient habitat ( $39 \times 10^6$ ). Likewise, the total CFU of fungi was also highest in native site ( $123.5 \times 10^3$ ). Correlation analysis indicated that, there was no relationship between the soil organic matter, total carbon on the soil bacterial and fungal population, however there was a strong relationship with the soil texture (Table 4). Surprisingly, the bacteria population had a negative relationship with the soil pH while the fungi had no relationship with the soil pH.

### AMF spores

The spore composition based on the morphology was different among the six sampling sites. Spores of different sizes and colors were present in all six sites. Highest morphological diversity was seen in the two recipient sites, and the lowest diversity was seen in recipient site 3. Spores of *Glomus* spp. were found in all the locations but were dominant in native site 3, and recipient site 2; spores of *Scutellospora* spp. were dominant in recipient site 1 and 3; and spores of *Gigaspora* spp. in native site 1 (Fig. 3).

## Discussion

Objective of this study was to analyze the role of soil rhizosphere, soil chemistry, and biology on the success of an invasive species in the recipient habitat compared to its native habitat. While our study was limited in the number of sampling sites and sampling frequency, our results show that there was a significant difference in the soil chemical, biological as well as physical characteristics in these two habitats, native and recipient. These soil characteristics can potentially, on their own or in combination with other habitat features, promote the extensive growth of exotic invasive plant in the recipient habitat compared to its native habitat.

Our results show that *L. microphyllum* has adapted to nutrient poor and highly acidic soils in its native habitat. In the recipient habitat, the plant can perform equally well in acidic soils and outcompete coexisting vegetation in slightly higher pH levels. Soil pH between 5.5 to 7.5 is known to provide the optimal growing conditions for this plant (Soti et al. 2015). The recipient sites in Florida, where *L. microphyllum* is invasive, had the optimum soil pH range. In addition, this pH range is also optimal for AMF which has been found to enhance the growth of *L. microphyllum* (Soti et al. 2014). In the native sites Australia, the soil pH was significantly lower than optimal for both the plant and AMF.

Along with strong acidic conditions, the native soils also had high concentration of Al, which is considered to be phytotoxic in strong acidic soils (Delhaize and Ryan 1995; Lidon and Barreiro 2002; Kochian et al. 2005). One of the sites in recipient habitat, where *L.*

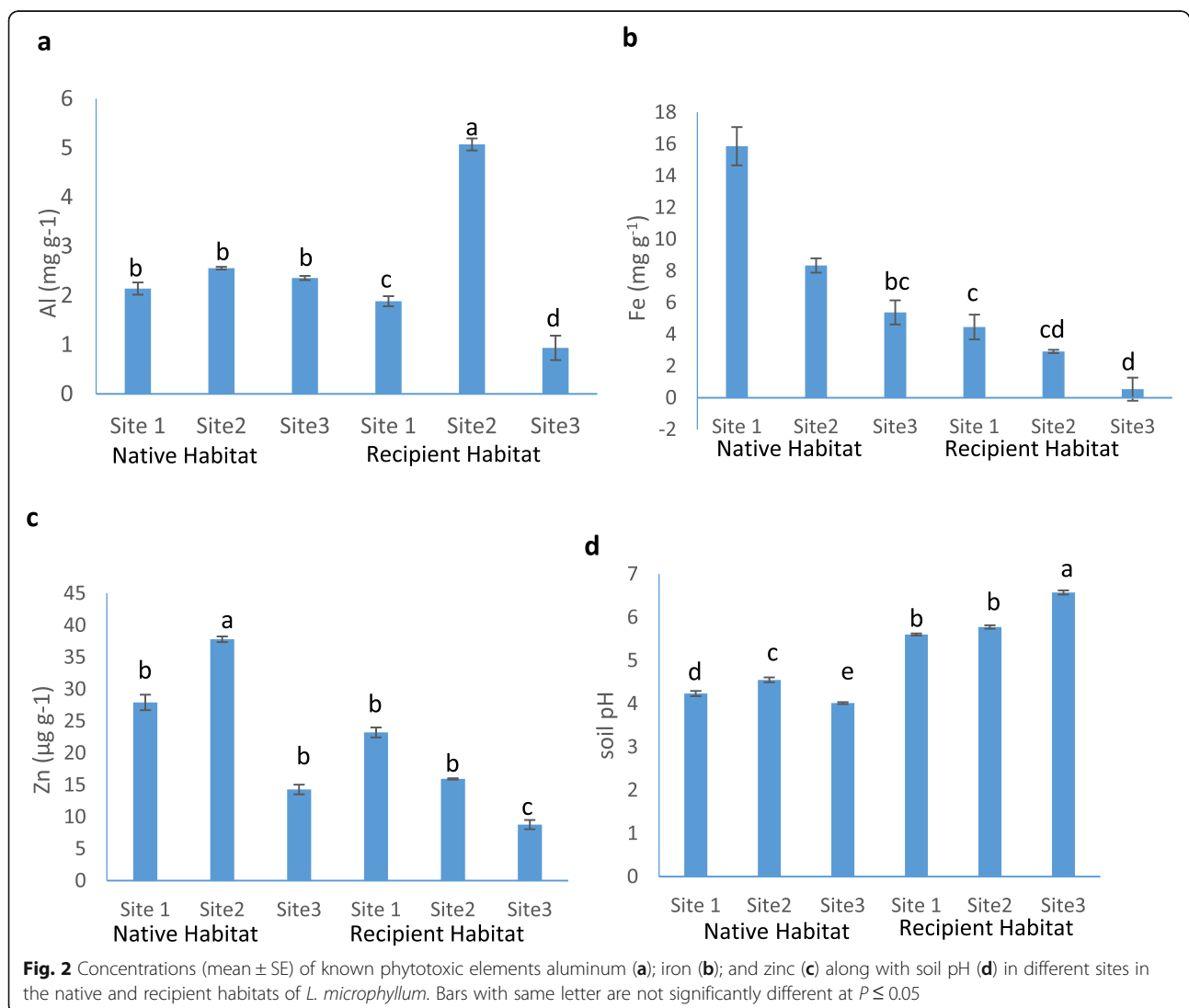
**Table 2** Soil texture and mean  $\pm$  standard error of the means of total colony forming units (counts  $\times 10^6$ ) of bacteria; (counts  $\times 10^3$ ) of fungi per gram of soil in the invaded and native sites

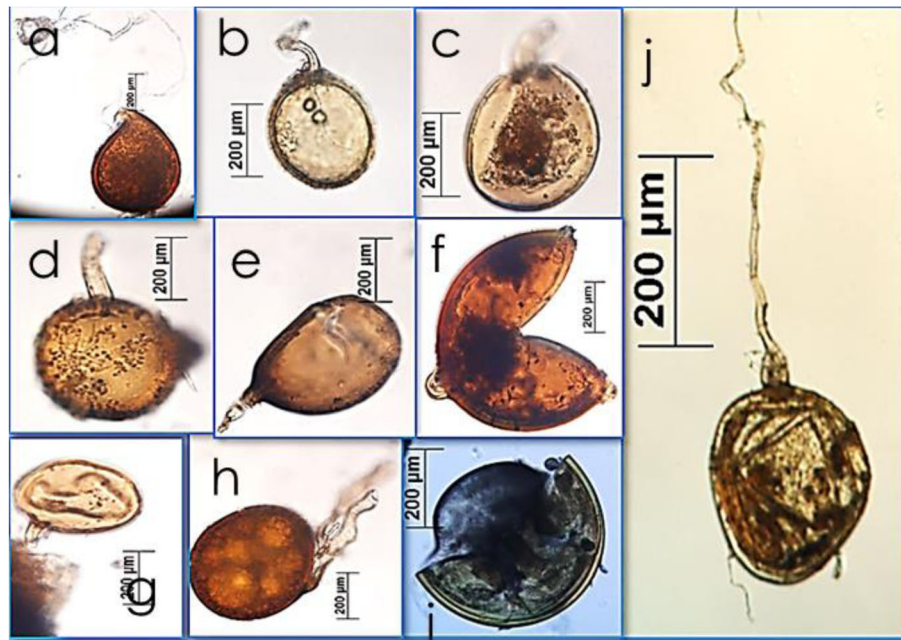
Sites	Soil Texture	Sand %	Silt %	Clay %	CFU of Bacteria $g^{-1}$ soil	CFU of Fungi $g^{-1}$ soil
Tree Tops Park, FL, US	Sandy loam	78	16.5	5.5	143.66 (13.00)	73.83 (12.27)
Central Florida, US	Clay	13	40	47	138.33 (10.78)	61.16 (9.82)
Jonathan Dickinson, FL, US	Sand	98	2	0	39 (4.28)	46.66 (7.76)
Daintree Ferry, Queensland, AU	Silt loam	25	57.5	17.5	288 (16.56)	123.5 (13.63)
Logan Reserve, Queensland, AU	Sandy clay loam	67.5	20	12.5	103.83 (10.12)	50.5 (8.75)
Nudgee, Queensland, AU	Loam	50	32.5	17.5	153.5 (6.15)	49.00 (4.60)

**Table 3** Mean (Std. Dev.) of the selected soil chemical characteristics in the native sites in Australia and invaded sites in Florida

Variable	Central Florida (FL)	Daintree (AU)	Jonathan Dickinson (FL)	Logan (AU)	Nudgee (AU)	Tree Tops (FL)
C %	4.03 (0.84)c	2.70 (0.44)c	3.02 (1.88) c	12.90 (2.43)b	4.28 (1.19)c	22.43 (4.15)a
Ca (mg g <sup>-1</sup> )	0.41 (0.13)b	0.47 (0.11)b	3.35 (1.77)b	0.43 (0.25)b	0.09 (0.01)b	17.21 (6.31)a
Cu (μg g <sup>-1</sup> )	8.42 (2.81)b	43.94 (12.31)a	1.84 (0.54)c	10.16 (0.57)b	6.74 (1.57)b	14.76 (4.02)b
K (mg g <sup>-1</sup> )	0.16 (0.10)b	1.77 (0.35)a	0.09 (0.05)b	1.49 (0.30)a	0.12 (0.02)b	0.18 (0.05)b
Mg (mg g <sup>-1</sup> )	0.20 (0.10)c	0.55 (0.18)b	0.21 (0.11)c	1.27 (0.17)a	0.25 (0.05)c	0.68 (0.27)b
Mn (mg g <sup>-1</sup> )	0.02 (0.01)b	0.27 (0.14)a	0.02 (0.01)b	0.02 (0.00)b	0.04 (0.00)b	0.05 (0.03)b
N %	0.20 (0.26)c	0.18 (0.03)c	0.49 (0.10)b	0.71 (0.04)b	0.26 (0.10)c	1.27 (0.17)a
OM %	8.65 (1.09)c	8.07 (2.89)c	4.32 (0.90)c	35.50 (7.04)b	11.45 (4.71)c	44.42 (2.71)a
P (mg g <sup>-1</sup> )	1.03 (0.25)a	0.91 (0.16)a	1.15 (0.09)a	1.16 (0.05)a	0.97 (0.06)a	1.22 (0.16)a

Means within rows followed by same letter are not significantly different at  $P \leq 0.05$





**Fig. 3** Dominant AMF morphotypes extracted from the rhizosphere soil of *L. microphyllum* in the both the native and recipient habitats. *Glomus* spp. spore (a, e, f); *Gigaspora* spp. (h, j); *Scutellospora* spp. (b, d, g, i)

*microphyllum* was growing over sand mine spoil, had the highest concentration of Al. However, this was not a restricting factor for *L. microphyllum* growth. Al toxicity is reported to cause alterations of physiological and biochemical process of plants and consequently in their productivity (Kochian 1995). Plant species differ in their Al tolerance, but *L. microphyllum* grown in Australian soil had lower biomass allocation to the belowground structures (rhizomes and roots) compared to the plants grown in Florida soils. When *L. microphyllum* escaped the highly acidic soil and clayey soil environment in the native habitat to the sandy and slightly alkaline soils in the new habitat, they could have acclimated to lowered investment cost in defense and reallocation of the resources to growth and reproduction, increasing their colonizing success.

Various soil and plant factors cause a significant influence in the soil microbial community, which have a fundamental role in nutrient cycling, plant growth and root health. It is widely reported that the rhizosphere community of different plant species growing in the same soil are distinct because of differences in root exudation

**Table 4** Correlation between CFUs of bacteria and fungi and soil parameters

	pH	OM%	N%	C%	Sand	Silt	Clay
CFU of Bacteria	-0.59*	-0.15	-0.09	-0.17	-0.71*	0.81*	0.41
CFU of Fungi	-0.21	-0.21	-0.15	-0.16	-0.64*	0.69*	0.44

\*Significant at  $P < 0.0001$

(Badri and Vivanco 2009; Brimecombe et al. 2000). Likewise, individual plant species can harbor different microbial communities in different soil types (Yang and Crowley 2000). A strong effect of soil texture on bacteria and fungi population was evident in this study. Native sites with the highest percentage of silt harbored highest CFUs of bacteria and fungi, while Jonathan Dickinson site in Florida with 98% sand had the lowest CFUs of bacteria and fungi. This kind of influence of soil texture on the structure of microbial population has been reported previously (Garbeva et al. 2004; Fang et al. 2005). An unexpected result was, the bacteria and fungi population remained uninfluenced by the soil organic matter, total C or soil pH which indicates a possible difference in the litter quality and secondary metabolites produced by the plant in its native range and invaded community and warrants further research.

Our results indicate that *L. microphyllum* had a symbiotic relationship with multiple species of AMF depending on the site conditions. Our two sites, Central Florida and Tree Tops which had higher diversities of spores are relatively disturbed sites compared to the other sites which had lower diversity of spores. This is an expected result and is in line with the Intermediate Disturbance Hypothesis (IDH) (Huston 1979), which suggests that a less disturbed healthy ecosystem has lower diversity of arbuscular mycorrhizal fungi. We found that *Glomus* sp., which is reported to be the dominant and most abundant genus of AMF, was present in all sites but

dominant in the native sites in Australia and one recipient site in Florida. Spores of *Scutellospora* sp. were dominant in the two-recipient site with slightly higher soil pH and sandier in texture. This study provides evidence that mycorrhizal fungi spore composition is different in the native and recipient habitats along with the soil characteristics, but an in-depth analysis with the use of molecular technique is necessary to identify the AMF species and their relationship with *L. microphyllum*. Future studies should focus on exploring the microbial community in the sites invaded by *L. microphyllum* and their role in its growth and competitive ability.

## Conclusions

Improved understanding of habitat characteristics of an invasive plant in its native and recipient range is important to understand whether the invaders have escaped hostile environmental conditions in their native range. Biogeographic variation in soil conditions could play an important role in the invasive success of nonnative plants in recipient habitats. Our results suggest that *L. microphyllum* can be growing poorly in its native range in Australia because of the soil toxic effects associated with soil acidity along with soil texture. In addition, while we were not able to determine the specific roles of the soil microbial community, difference in the microbial populations and types in the two regions could in part explain the extensive growth of *L. microphyllum* in the recipient habitats in south Florida. Thus, land managers should focus on soil restoration to manage the extensive growth of invasive species and increase the competitive ability of native species.

## Abbreviations

Al: Aluminum; AMF: Arbuscular mycorrhizal fungi; C: Carbon; Ca: Calcium; CFU: Colony forming unit; DI: Deionized; Fe: Iron; ICP-MS: Inductively coupled plasma mass spectrometry; Mg: Magnesium; Mn: Manganese; N: Nitrogen; P: Phosphorus; USDA-ARS: United States Department of Agriculture - Agricultural Research Service; Zn: Zinc

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## Authors' contributions

P. Soti: Designed the experiment, collected and analyzed the samples. P. Soti also, analyzed the data and wrote the manuscript. M. Purcell: Assisted in experimental design, collected the samples in Australia, and assisted in writing the manuscript. K. Jayachandran: Assisted in experimental design, provided resources, and assisted in writing the manuscript. The author(s) read and approved the final manuscript.

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## Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Ethics approval and consent to participate

Not applicable.

## Consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

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